Multilocus Genetic Divergence Within and Among Three Species of *Timema* Walking Sticks

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Patrik Nosil – Department of Ecology and Evolutionary Biology Barbara Demmig-Adams – Department of Ecology and Evolutionary Biology Andrew Martin – Department of Ecology and Evolutionary Biology Matthew Wilsey-Cleveland – Program for Writing and Rhetoric **Abstract:** The theory of ecological speciation suggests that adaptation to different habitats promotes the formation of new species (speciation), but debates persist about the extent to which this can occur when populations are in geographic contact and exchange genes (i.e. gene flow). Multilocus genetic analysis (using multiple genes) can inform this debate by comparing levels of genetic divergence between taxa (groups such as species) at different points in the speciation process and with differing geographic arrangements. For example, taxa undergoing less gene flow are predicted to exhibit stronger genetic divergence than taxa undergoing more gene flow. Furthermore, multilocus data can be used to test for genealogical discordance—a scenario where different genes tell different evolutionary histories, due to being differentially affected by evolutionary processes such as selection, mutation, and gene flow. The present study uses multilocus genetic data (four nuclear genes and a mitochondrial gene) to compare the genetic structure of six populations of herbivorous *Timema* walking-sticks with different geographic relationships: two diverging host-associated ecotypes (phenotypically and ecologically different groups) of T. cristinae in parapatry (geographic contact) and allopatry (geographic separation), and two closely related species, *T. californicum* and *T. poppensis*, in parapatry and allopatry. As expected, we observe that species are generally more genetically differentiated than ecotypes. Interestingly, however, allopatric populations are not always significantly more differentiated than parapatric populations. Also, genealogical discordance was observed. Mitochondrial differentiation among allopatric *T. cristinae* population was stronger than nuclear differentiation. In contrast, mitochondrial differentiation between the T. californicum and T. poppensis was weak, whereas nuclear genes were differentiated to the point of reciprocal monophyly, indicative of genetically distinct species. While these data

strongly suggest differing levels of gene flow among genes and taxa, further analyses using coalescent-based models should be done to estimate gene flow specifically.

Glossary

- <u>Ecological Speciation</u> the process in which reproductive isolation evolves between populations through ecologically based divergent selection
- <u>Ecologically Based Divergent Selection</u> selection arising from environmental differences and/or ecological interactions, which act in contrasting directions on two populations
- <u>Reproductive Isolation</u> a reduction or lack of genetic exchange (gene flow) between populations
- <u>Gene Flow</u> the transfer of genes or alleles from one population to another population
- <u>Ecotype</u> phenotypically and ecologically different groups or populations within the same species
- <u>Multilocus Genetic Analysis</u> analysis utilizing genetic sequence data at multiple different locations across the genome
- <u>Speciation Genes</u> genes whose divergence made a significant contribution the evolution of reproductive isolation between populations or any gene that reduces hybrid fitness

<u>Neutral Genes</u> – genes that have no significant role in the speciation process

- <u>Adaptive Genetic Divergence</u> the process of genetic differentiation between two populations arising from different sets of environmental conditions to become more suited for
- <u>Assortative Mating</u> mating of individuals which share more common traits (e.g. individuals of the same ecotype) than likely from random mating
- <u>Amplified Fragment Length Polymorphism (AFLP)</u> a molecular method for identifying DNA polymorphisms, used to detect loci that exceed neutral expectations (speciation genes)
- <u>Reciprocal Monophyly</u> mutually exclusive groups of individuals, characteristic of distinct species

Introduction

There are historical and current debates regarding the underlying mechanisms and geographic modes of speciation (Coyne & Orr, 2004). Some assert that divergence can only proceed if gene flow (intermixing of genes between populations) is absent or very limited, while others claim that divergence can indeed take place in the face of gene flow. Furthermore, it remains unclear what the implications of gene flow are for the rate of speciation or for how far the process of divergence proceeds (Nosil, Harmon, & Seehausen, 2009; Via, 2001). Another more rudimentary debate is simply how to define a species. This issue is known as the "species problem", comprised of numerous working definitions for a species (Hey, 2006a). For the purposes of this project, I will use Ernst Mayr's (1942) biological species concept that a species is an interbreeding group of individuals, reproductively isolated from other groups. Here, speciation is most simply the evolutionary process of forming new "biological" species, trending toward reduced gene flow and increased genetic divergence between taxa as speciation unfolds. In this context, speciation may refer to the evolution from one species into another, or the divergence of one ancestral species into two or more new species (Dobzhansky, 1940). Many have argued that the species problem exists, at least in part, because speciation occurs along a continuum of divergence rather than as a discrete event (Mallet, 2007; Mallet, Beltran, Neukirchen, & Linares, 2007). Over time, differentiation gradually accumulates until two populations become strongly or fully reproductively isolated species. The extended and continuous nature of the speciation process thus makes it difficult to define the exact point at which two diverging populations have become distinct species (Mallet, 2007).

The geographic arrangement of populations is also important when discussing the processes of evolutionary divergence. Speciation was originally thought to require populations

in allopatry (geographic isolation). Mayr, a large contributor to this field, claimed that speciation is unlikely to happen under conditions other than strict allopatry (Futuyma & Mayer, 1980; Mayr, 1942, 1947, 1954). This view implies that geographic isolation is required to act as the barrier to gene flow and allows for the build-up of genetic differences between populations, which incidentally cause reproductive isolation. Under this hypothesis, isolated subpopulations adapt to different environmental conditions and experience random genetic mutation and genetic drift (random change in allele frequencies), resulting in genetic differentiation and thus the eventual formation of new species over time. Intuitively, this makes sense, especially under the assumption that gene flow would break down the process of genetic divergence and speciation (Mayr, 1963; Rice & Hostert, 1993; Wright, 1931). However, with new technology focused on generating large genetic datasets, and new theoretical and conceptual arguments being made, there have been many challenges to the view of strict allopatric speciation (Futuyma & Mayer, 1980; Nosil, 2008; Rice & Hostert, 1993; White, 1978). It is now clear that speciation happens (at least some of the time) in sympatry, where populations occur in the same geographic area, and parapatry, where adjacent populations share a marginal area of overlap (Rice & Hostert, 1993; Rundle & Nosil, 2005). In nonallopatric models, barriers to gene flow no longer need arise from geographic constraints but rather are initiated through diversifying evolutionary processes (Nosil & Rundle, 2009). For example, reproductive isolation might evolve as a by-product of adaptive genetic divergence, stemming from ecological differences between populations and subsequent ecologically based divergent selection (i.e. ecological speciation) (Rundle & Nosil, 2005), or via the process of "reinforcement", where a reduction in hybrid offspring fitness causes selection against hybridization and for assortative mating (Rice & Hostert, 1993; Rundle & Nosil, 2005).

With a growing understanding of the complex nature of speciation, many historical assumptions have come into question; particularly, that gene flow must be absent during the process of speciation. More specifically, speciation scenarios where gene flow may be present involve sympatric or parapatric populations that undergo ecological speciation and/or reinforcement. Although the end of speciation is marked by complete reproductive isolation (no genetic exchange), it does not imply that gene flow must not have occurred at some point during divergence. In fact, the possibility of divergence with gene flow has been shown in a number of theoretical scenarios (Bolnick & Fitzpatrick, 2007; Nosil, 2008), but the number of empirical examples of speciation with gene flow remains limited (Emelianov, Marec, & Mallet, 2004; Faure, Jollivet, Tanguy, Bonhomme, & Bierne, 2009; Niemiller, Fitzpatrick, & Miller, 2008). Recent advances in technology have increased the ease of acquisition of genetic sequence data, and have allowed for the use of such data for population level analyses. Because genetic information provides the best insight into the evolutionary histories of extant taxa and because of the statistical power gained from large sample sizes, population level genetic data are necessary to make accurate inferences about evolutionary divergence and gene flow. Despite substantial improvements, evolutionary geneticists are still troubled with reconstructing the genetic histories that involve gene flow during divergence, leaving the role of gene flow in speciation not yet fully understood.

Gene Flow and Population Differentiation:

Gene flow, selection, and mutation are three key processes involved in genetic differentiation and subsequent speciation (Coyne & Orr, 2004). Mutation generates new genetic variants, which can then be acted upon by selection or genetic drift. These latter processes work together to establish genetic differentiation between diverging species, while

gene flow may break down or reinforce divergence (Tajima, 1983). Genetic differentiation itself can be assessed through the use of F_{ST}, the inbreeding coefficient. An F_{ST} value of 0 (or values not significantly different from 0) indicates little to no genetic differentiation and an F_{ST} value of 1 reflects complete genetic differentiation (i.e. fixed differences) (Wright, 1965). For example, low F_{ST} values might reflect populations experiencing strong gene flow, while high F_{ST} values might suggest that little or no gene flow is occurring. However, a major difficulty here is disentangling the effects of gene flow versus divergence time on levels of differentiation. For example, low F_{ST} values might arise from high gene flow, recent divergence (i.e. insufficient time to accumulate differentiation), or a combination of these processes (Hey, 2006b).

Genetic Differentiation and Genealogical Discordance:

In addition to varying among different pairs of populations, levels of genetic differentiation can vary among regions of an organism's genome. By using multilocus genetic data, one can compare the amount of differentiation at each locus. When different loci (or genes) show different amounts of genetic differentiation, and thus illustrate different evolutionary histories, it is known as genealogical discordance (Degnan & Rosenberg, 2009). Variation among F_{ST} estimates between different loci can provide useful information about how selection and gene flow differentially affect gene regions (Beaumont, 2005). For example, genes subject to divergent selection are expected to be more differentiated than other genes, because selection is essentially pulling apart allele frequencies between populations at selected loci and preventing alleles at these loci from moving between populations.

While mutation, selection, and gene flow all influence genetic differentiation, they are also the root causes of genealogical discordance. Mutation, a stochastic process, does not occur at equal rates across the genome (Neigel & Avise, 1986). Selection then plays a major role in the differential rates at which mutations (or genetic differentiation) accumulate in a population. For example, if a mutation occurs in a gene that is directly involved in establishing reproductive isolation (speciation gene), selection will remove that mutation from the population if it proves to be disadvantageous, while mutations in genes not involved in speciation (neutral genes) may persist. Also, genes do not flow at equal rates between populations (Wang, Wakeley, & Hey, 1997). Similar to the accumulation of mutation, divergent selection plays a critical role in determining which genes will or will not "flow". For speciation genes, some alleles will be advantageous for one ecotype and disadvantageous for another ecotype. These alleles will be selected differentially and will therefore "flow" less readily when introgression (mating between diverging populations) occurs (Wang, et al., 1997). Furthermore, while neutral genes that are not physically linked to genes under divergent selection may flow readily between populations, gene flow at neutral loci that are linked to speciation genes will be resisted (Barton, 1998; Smith & Haigh, 1974). For these reasons, it is common for different genes to reveal remarkably different evolutionary histories, which is shown as highly variable levels of genetic divergence between populations across different loci. One such example of genealogical discordance is the contrast between mitochondrial genes and nuclear genes. Mitochondrial DNA is estimated to evolve 5 to 10 times faster than nuclear DNA (Brown, George, & Wilson, 1979). There are many potential explanations for this pattern including: (1) mitochondrial DNA replicates more frequently than nuclear DNA and therefore has a greater likelihood of mutating, (2) mitochondrial DNA is only inherited maternally,

resulting in a smaller effective population size and (3) mitochondrial DNA does not undergo recombination events, causing mutations to accumulate more quickly in a population (Brown, et al., 1979; Dopman, Perez, Bogdnowicz, & Harrison, 2005).

In sum, although genealogical discordance is expected to occur even in the absence of gene flow, simply due to the stochasticity of genetic drift, discordance is expected to be greater when gene flow occurs, because gene flow is affected at some regions (e.g. speciation genes or divergently selected regions) more than others (e.g. neutral regions) (Hey, 2006b).

Study System: Timema walking sticks

Timema walking sticks are wingless, herbivorous insects found primarily in the chaparral of western North America (Vickery, 1993). The genus *Timema* is comprised of approximately 20 different species of walking stick and is estimated to have emerged 20 million years ago in conjunction with the spread of the chaparral biome (Crespi & Sandoval, 2000; Sandoval, Carmean, & Crespi, 1998). *Timema* are highly cryptic insects, resting on leaves and stems during the day and feeding at night (Crespi & Sandoval, 2000). A large body of research supports the conclusion that ecological speciation is occurring in different populations of the species *Timema cristinae* (Nosil, 2007). Notably, *T. cristinae* is in the early stages of ecological speciation and found as two different host-plant associated "ecotypes", or phenotypically and ecologically different groups. These ecotypes are distinguished by numerous morphological differences, with one of the most pronounced being the presence or absence of a dorsal stripe (along the back of the insect). The stripe is common on the host Adenostoma fasciculatum, a plant with thin needle-like leaves. In contrast, the unstriped ecotype is common on *Ceanothus spinosus*, a broad-leaved plant (Nosil, 2007). It is clear from past experimental and genotypic (e.g. amplified fragment length polymorphisms [AFLP]) data

that these two ecotypes are undergoing ecological speciation in the presence of gene flow though host-plant adaptation and ecologically based divergent selection (Nosil, 2007; Nosil, Crespi, & Sandoval, 2003; Nosil & Rundle, 2009). However, multilocus DNA sequences data have not yet been analyzed to make, for example, independent estimates of divergence time versus gene flow (Nosil, et al., 2003). Other members of the genus, namely *Timema poppensis* and *Timema californicum*, have unclear phylogenetic histories (Law & Crespi, 2002). Based on sequence divergence in the mitochondrial gene cytochrome oxidase 1 (CO1), a gene commonly used for inferring insect phylogenies, the latter two "species" appear to be closely related or even the *same species*, but are phenotypically highly distinct (Law & Crespi, 2002). Since genealogical discordance can provide information on gene flow and divergence time, further work on these taxa utilizing multiple loci is necessary.

Hypotheses and Questions:

I predict that the species pairs will exhibit greater genetic divergence than the ecotype pairs, given that species are further along the speciation continuum than ecotypes. Also, I predict that estimates of F_{ST} will differ among genes, especially so between mitochondrial genes and nuclear genes. Furthermore, it is likely that mitochondrial DNA will show a different evolutionary history than nuclear DNA, given what is known about genealogical discordance and mitochondrial evolution. Also, I will examine the role of geographic arrangement during speciation and more specifically, see if geographic arrangement is playing a role in the differentiation of *T. cristinae* or if ecologically based divergent selection is driving factor.

Methods

Study Populations and Sampling:

In order to make comparisons of genetic divergence for varying degrees of speciation (how much divergence has occurred), we compared divergence between ecotypes of *Timema* cristinae to divergence between the species pair, Timema californicum and Timema poppensis. To also allow for the examination of the effects of geographic arrangement, we examined two adjacent parapatric populations ("parapatric pair" hereafter) and two geographically separated populations ("allopatric pair" hereafter) for both ecotype and species level comparisons (see Table 1). The recent and incomplete ecological speciation of T. cristinae ecotypes could represent a model of recent divergence, while T. poppensis and T. californicum could fit a model of more ancient divergence. By using both parapatic and allopatric populations for comparisons we are able to assess the level of genetic divergence given these geographic scenarios. Previous experimental and genetic research indicates that *Timema* do not disperse over long distances, resulting in limited interbreeding between geographically separated populations (Crespi & Sandoval, 2000; Nosil, et al., 2003). Thus, these allopatric populations can be considered genetically isolated from their respective parapatric counterparts, making them a sufficient comparison with the parapatric population pairs to discern genetic divergence due to isolation and assess the impacts of gene flow at multiple loci.

Approximately 25 specimens from each of the six populations were collected during their breeding season, between March and June, in 2010. The parapatric ecotypes of *T. cristinae* were collected from the populations HVC and HVA (N34 29.309 W119 47.180 and N34 29.305 W119 47.191 respectively) and the allopatric *Adenostoma* ecotype was collected at LA (N34 30.464 W119 47.694). Individuals from parapatric populations of *T. poppensis*

(PSM1RW) and *T. californicum* (CSMQ) were collected along Summit Road (N37 133.43 W122 05.271) and individuals from the allopatric population of *T. poppensis* were collected at PTBDF (N38 37.100 W123 17.322). Once collected, all samples were stored in 100 percent ethanol, and shipped to the University of Colorado at Boulder to be stored at -40°C.



Figure 1. Study Design: Ecotype and species comparisons by populations based on geographic arrangement. The ecotypes of *Timema cristinae* are represented by three populations, two on the plant-host *Adenostoma* (HVA and LA) and one on *Ceanothus* (HVC). *Timema poppensis* is represented by two populations (PSM1RW and PTBRW) and *Timema californicum* is represented by one popuations (CSMQ).

Molecular Methods:

DNA was extracted from *Timema* legs using Quiagen DNEasy Blood and Tissue Kits, following the DNEasy protocol (Quiagen, Valencia, CA). Depending on the size and quantity of the legs available, samples were eluted to a final volume of 100 - 150 µl, so as to obtain optimal DNA concentrations. All extractions were checked using a NanoDrop (micro-sample quantification instrument for nucleic acid concentration) (Thermo Scientific) to determine DNA concentration. Extraction products were kept frozen at -20°C for later use.

Nuclear and mitochondrial primers (short nucleic acid strands that designate the beginning and end of the amplification region) were developed based on mRNA sequences of T. cristinae obtained from 454 pyrosequencing of normalized cDNA libraries (P. Nosil, unpublished). Four nuclear loci (N1, N2, N3 and N4) and one mitochondrial locus, cytochrome oxidase 1 (CO1), were amplified in the six populations of *Timema* using polymerase chain reaction (PCR) techniques with the following reaction conditions: 5µl of 5x buffer, 0.25µl dNTPs (10mM), 2.5µM MgCl₂, 0.2µM forward and reverse primers, 0.25µl Taq polymerase; 23µl of this master mix and 2µl template DNA were used per reaction. All nuclear genes were amplified with a PCR cycling profile of 95°C for 3 minutes followed by 35 cycles of 95°C for 1 minute, 50°C for 40 seconds and 72°C for 45 seconds, finished with 72°C for 7 minutes. Mitochondrial CO1 was amplified with a PCR cycling profile of 94°C for 5 minutes followed by 30 cycles of 94°C for 1 minute, 43°C for 1 minute and 72°C for 50 seconds and finished with 72°C for 5 minutes. All PCR amplifications were executed using Mastercycler pro S (Eppendorf). The amplified products were run on 2% agarose gels, in combination with cybergreen and loading dye using standard electrophoresis techniques to check for successful amplification via ultra-violet imaging.

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Locus Name	L Pr	rimer Sequences (5' - 3')
Nuclear		
N1	425	F-ATCCTGGAATTCACGCACTTAC
111	425	R-CTTACCCTTCTCCAAAATGTCG
N2	421	F-AAATTGCATTATCCAGGTCAGC
112	721	R-GTTTATTCAGAAGCCCGTTGTC
NB	380	F-AGCCTCTATTGATCGTCGAAAC
NJ	500	R-TCAATTTCACGATTTTCACTCG
N4	455	F-AAAGCACCCAAGAAGAATGTTG
11-1	455	R-CAGCATGGTTGGTTCATAAGTG
Mitochondrial		
	64.0	F-GGTCAACAAATCATAAAGATATTGG
01	612	R-TAAACTTCAGGGTGCCAAAAAATCA

Table 1. Locus details and PCR conditions.

DNA sequencing was performed using traditional Sanger sequencing methods at Arizona State University (Tempe, AZ) (Sanger, Nicklen, & Coulson, 1977). Forward and reverse sequences were assembled into contigs, aligned and edited using the program Geneious 5.3.4 (Drummond, et al., 2010). Ambiguous, heterozygous loci in nuclear genes were parsed using Phase 2.1.1, which uses Bayesian methods for haplotype reconstruction from genetic sequence data (Stephens & Donnelly, 2003; Stephens, Smith, & Donnelly, 2001). It is important to note here that genetic sequence data for CO1 in the allopatric population of *T. poppensis* was unobtainable. This is likely due to the primers being developed based on *T. cristinae* mRNA sequences and potential mutations in *T. poppensis* at the priming region causing irregular or inconsistent amplification. This problem was also observed for 13 individuals from the *T. poppensis* population (PSM1RW) resulting in a markedly reduced sample size (n=15) for CO1.

Population Statistics:

Within population summary statistics, including sample size (n) the numbers of haplotypes (h), genetic diversity, percent of polymorphic sites (%S) and Tajima's D, a test of selective neutrality, were all calculated using Arliquin 3.5.1.2 (Excoffier & Lischer, 2010) (*see Table 2*). Tajima's D indicates whether a gene is evolving via random neutral processes (i.e. drift) or via nonrandom mechanisms (i.e. divergent selection). Values greater than two or less than minus two roughly indicate differentiation via nonrandom processes such as divergent selection or other non-random forms of differentiation (Tajima, 1989). Pairwise F_{ST} values were estimated using the same software to provide information about the degree of genetic differentiation between populations (Weir & Cockerham, 1984).

Phylogenetic Analyses:

A priori neighbor-joining trees were constructed for each locus using all populations of *Timema*. For the construction of these unrooted trees, the resampling method of bootstrapping with 1000 replicates was utilized in the program MEGA 5.0 (Tamura, et al., 2011). For all trees (e.g. at every locus) *T. cristinae* was reciprocally monophyletic from *T. poppensis/T. californicum* with 100% confidence, justifying subsequent phylogenetic analyses within in each clade (*T. cristinae* clade and *T. poppensis/T. californicum* clade) separately. In these subsequent analyses, *T. cristinae* was used to root the "*T. poppensis/T.californicum* trees" and *T. poppensis* was used to root "*T. cristinae* trees". The rooted gene trees for ecotype and species comparisons were constructed for all four nuclear loci and mitochondrial CO1 using Bayesian Markov chain Monte Carlo (MCMC) methods. These phylogenetic analyses were conducted to reconstruct phylogenies using MrBayes v3.0b4 (Ronquist & Huelsenbeck, 2003), implementing the GTR + I + Γ model of evolution.

Two simultaneous runs of four MCMC analyses, consisting of one cold and three incrementally heated chains, were initiated with random trees for a total of 5.0×10^5 generations (sampling every 100 generations). The first 2.5×10^5 generations from each run were discarded as burn-in.

Population Structure:

The program Structure 2.3.3 was used to visualize the genetic structure of the six populations. An admixture model of ancestry was used so as to assume each individual consists of DNA from any of the defined populations. Additionally, a model of independent allele frequencies was assumed in order to allow for different populations to have different allelic frequencies (Pritchard, Stephens, & Donnelly, 2000). The parameter (K) indicates the number of predefined genetic clusters. For these analyses K = 3 and K = 6 were used due to the biological justifications: three species and six populations, respectively. Analyses were executed using 100,000 replicates and a 10,000 burn-in.

Results

Within Population Genetic Diversity:

Within population summary statistics reveal significant genetic variability within each population, and also some differences in the level of variability among the different gene loci (*see Table 2*). For example, percent polymorphic sites (number of variable sites over sequence length) were highest in mitochondrial locus CO1 and lowest in nuclear loci N3 and N4. Also, the number of haplotypes for each locus and population were respectively higher in nuclear genes (e.g. N1 and N2) than the mitochondrial gene. Also, Tajima's D values generally fell within the range of expectation for neutrally evolving loci (i.e. between two and minus two, *Table 2*).

Locus	Population	n	h	Genetic Diversity	%S	Tajima's D
Nuclear						
235	HVC	50	26	0.93 ± 0.02	4.71	-1.24
	HVA	52	24	0.94 ± 0.02	4.24	-0.94
	LA	46	17	0.93 ± 0.02	5.41	-1.28
	CSMQ	50	5	0.65 ± 0.04	0.94	1.79
	PSM1RW	54	2	0.11 ± 0.06	0.47	-0.90
	PTBDF	48	1	0.00 ± 0.00	0.00	0.00
9172	HVC	44	8	0.63 ± 0.08	2.61	-1.48
	HVA	50	9	0.67 ± 0.07	2.85	-1.14
	LA	44	8	0.76 ± 0.05	2.38	-1.10
	CSMQ	46	4	0.24 ± 0.08	0.95	-1.45
	PSM1RW	54	2	0.07 ± 0.05	0.24	-0.88
	PTBDF	48	1	0.00 ± 0.00	0.00	0.00
10854	HVC	48	3	0.20 ± 0.07	0.53	-1.00
	HVA	52	3	0.21 ± 0.07	0.53	-0.89
	LA	44	3	0.28 ± 0.08	0.53	-0.66
	CSMQ	50	2	0.18 ± 0.07	0.26	-0.24
	PSM1RW	52	2	0.27 ± 0.07	0.26	0.27
	PTBDF	48	2	0.51 ± 0.02	0.53	2.27
11144	HVC	48	1	0.00 ± 0.00	0.00	0.00
	HVA	52	2	0.04 ± 0.04	0.22	-1.10
	LA	42	2	0.09 ± 0.06	0.22	-0.85
	CSMQ	48	2	0.04 ± 0.04	0.88	-1.87
	PSM1RW	48	3	0.45 ± 0.06	0.44	1.64
	PTBDF	44	1	0.00 ± 0.00	0.00	0.00
Mitochondrial						
CO1	HVC	25	7	0.81 ± 0.05	8.006536	1.44
	HVA	26	7	0.85 ± 0.03	8.006536	2.10
	LA	21	5	0.64 ± 0.10	7.51634	0.51
	CSMQ	25	8	0.71 ± 0.09	1.960784	-0.06
	PSM1RW	15	4	0.72 ± 0.08	1.470588	1.54
	PTBDF	N/A	N/A	N/A	N/A	N/A

Table 2. Within population descriptive statistics including sample size (n), number of haplotypes (h), genetic diversity, percent polymorphic sites (%S) and Taiima's D.

Between Population Genetic Differentiation:

Estimates of population differentiation are presented in Table 3. For nuclear genes, F_{ST} values (indicating the level of genetic differentiation) between both the parapatric pair and allopatric pair of *T. cristinae* illustrate no significant differentiation. However, for

mitochondrial CO1, the F_{ST} values indicate significant differentiation between allopatric *T. cristinae* populations (0.34), but no differentiation between parapatric populations (not significantly different from 0). In contrast to what was observed between ecotypes of *T. cristinae*, F_{ST} estimates between the species pair (*T. californicum* and *T. poppensis*) illustrate significant and very strong genetic differentiation at all nuclear loci between both the parapatric and the allopatric pairs (ranging from 0.72 to 0.93). In contrast to nuclear differentiation between species, mitochondrial CO1 was weakly differentiated between *T. californicum* and *T. poppensis* (*see Table 3*).

Table 3. F_{ST}: The inbreeding coefficent, a measure of genetic divergence.
 * indicates significance (e.g. significantly different from zero) (p-value < 0.05)

Population Pair	N1	N2	N3	N4	CO1
HVC x HVA	0.016	-0.008	-0.004	-0.002	0.03
HVC x LA	-0.013	0.026	0.004	0.029	0.343*
CSMQ x PSM1RW	0.924*	0.904*	0.884*	0.896*	0.331*
CSMQ x PTBDF	0.929*	0.917*	0.717*	0.982*	N/A

Gene Trees:

Consistent with our findings from F_{ST}, nuclear gene trees show no significant differentiation among all *T. cristinae* populations, illustrated by a comb-like tree structure at all nuclear loci (*see Figures 2-5*). In contrast, there was some differentiation among conspecific populations of *T. cristinae* in the mitochondrial gene tree, grouping most of the allopatric population (LA) together, but with both parapatric populations and LA being nonetheless mixed, suggesting the early stages of genetic divergence at CO1 in allopatry.

Nuclear gene trees for the *T. poppensis/T. califronicum* species pair comparison show complete differentiation, or species-level reciprocal monophyly, for all nuclear loci *(see Figures 2-5).* In contrast, the mitochondrial gene tree for CO1 suggests a mixed relationship between the species, *T. californicum* and *T. poppensis*, consistent with the findings of Law and Crespi, 2002 *(see Figure 6).*

HVC 1					OUTGROUP
HVC_2	HVC 3				
HVC_4 HVC_5	1140_0				
HVC_6 HVC_7					
 HVC 9		HVC_8			
HVC_10 HVC_11					
 HVC_12 HVC_13					
- HVC_14	HVC_15				
 HVC_16 HVC_17					
 HVC_18 HVC_19					
 HVC_20 HVC_21					
 HVC_22 HVC_23					
 HVC_24 HVC_25					
 HVA_2 HVA_3					
 HVA_4					
 HVA 6					
HVA_8 HVA_9					
HVA_10					
HVA_12 HVA_13					
 HVA_14 HVA_15					
HVA_16 HVA_17					
HVA_18 HVA_19					
HVA_20 HVA_21					
HVA_22 HVA_23					
 HVA_24 HVA_25					
HVA_26					
LA_2 LA_3					
LA_4 LA_5					
LA_6 LA_7					
 LA_0 LA_9					
 LA 10					
 LA_13					
LA_14 LA_15					
LA 18					
LA_19					
LA_21	LA 22				
 - LA_23	-				
			0.2		
			0.2		
				CSMQ_1	OUTGROUP
			-	CSMQ_1 CSMQ_2 CSMQ_3	OUTGROUP
				CSMQ_1 CSMQ_2 CSMQ_3 CSMQ_4 CSMQ_4	OUTGROUP
				CSMO_1 CSMO_2 CSMO_2 CSMO_3 CSMO_4 CSMO_6 CSMO_6 CSMO_7	OUTGROUP
				CSMO_1 CSMO_2 CSMO_2 CSMO_5 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_10	OUTGROUP
				CSMO_1 CSMO_2 CSMO_3 CSMO_3 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_7 CSMO_7 CSMO_7 CSMO_1 CS	OUTGROUP
				CSMQ_1 CSMQ_2 CSMQ_3 CSMQ_4 CSMQ_6 CSMQ_6 CSMQ_7 CSMQ_9 CSMQ_9 CSMQ_10 CSMQ_112 CSMQ_13 CSMQ_13 CSMQ_14	OUTGROUP
				CSM0 1 CSM0 3 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 8 CSM0 8 CSM0 10 CSM0 11 CSM0 13 CSM0 13 CSM0 13 CSM0 16	OUTGROUP
				CSMO 1 CSMO 3 CSMO 3 CSMO 5 CSMO 6 CSMO 6 CSMO 7 CSMO 7 CSMO 7 CSMO 10 CSMO 10 CSMO 11 CSMO 14 CSMO 16 CSMO 16 CSMO 16 CSMO 18	OUTGROUP
				CSMQ_1 CSMQ_3 CSMQ_5 CSMQ_5 CSMQ_6 CSMQ_6 CSMQ_6 CSMQ_7 CSMQ_10 CSMQ_10 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_15 CSMQ_16 CSMQ_15 CSMQ_16 CSMQ_1	OUTGROUP
				CSMO_1 CSMO_3 CSMO_5 CSMO_6 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_10 CSMO_11 CSMO_11 CSMO_11 CSMO_14 CSMO_15 CSMO_16 CSMO_16 CSMO_16 CSMO_19 CSMO_12 CSMO_12 CSMO_12 CSMO_12 CSMO_12 CSMO_12 CSMO_12 CSMO_12 CSMO_12 CSMO_22 CS	OUTGROUP
				CSMO_1 CSMO_3 CSMO_5 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_10 CSMO_11 CSMO_11 CSMO_12 CSMO_12 CSMO_12 CSMO_14 CSMO_15 CSMO_16 CSMO_17 CSMO_17 CSMO_17 CSMO_12 CSMO_12 CSMO_12 CSMO_22 CSMO_24 CSMO_24	OUTGROUP
			PSMIRW_1	CSM0 1 CSM0 3 CSM0 4 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 8 CSM0 8 CSM0 8 CSM0 10 CSM0 11 CSM0 13 CSM0 13 CSM0 13 CSM0 14 CSM0 13 CSM0 14 CSM0 12 CSM0 12 CSM0 20 CSM0 21 CSM0 21 CSM0 23 CSM0 25	OUTGROUP
			PSM1RW_1 PSM1RW_2 PSM1RW_3	CSM0 1 CSM0 3 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 5	CSMO_1 CSMO_3 CSMO_3 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_7 CSMO_10 CSMO_11 CSMO_11 CSMO_11 CSMO_13 CSMO_13 CSMO_14 CSMO_14 CSMO_14 CSMO_16 CSMO_16 CSMO_16 CSMO_16 CSMO_12 CSMO_12 CSMO_25 CSMO_22 CSMO_22 CSMO_22	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 4 PSM1RW 5 PSM1RW 5 PSM1RW 5	CSMQ_1 CSMQ_3 CSMQ_5 CSMQ_6 CSMQ_6 CSMQ_6 CSMQ_6 CSMQ_7 CSMQ_10 CSMQ_10 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_11 CSMQ_12 CSMQ_20 CSMQ_20 CSMQ_21 CSMQ_21 CSMQ_21 CSMQ_22 CSMQ_22 CSMQ_22	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 2 PSM1RW 5 PSM1RW 5 PSM1RW 6 PSM1RW 6 PSM1RW 6 PSM1RW 7 PSM1RW 7 PSM1RW 9 PSM1RW 9	CSM0 1 CSM0 3 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CSM0 10 CSM0 11 CSM0 11 CSM0 11 CSM0 11 CSM0 13 CSM0 14 CSM0 16 CSM0 16 CSM0 16 CSM0 20 CSM0 20 CSM0 21 CSM0 21 CSM0 21 CSM0 22 CSM0 22	OUTGROUP
			PSM1RW_1 PSM1RW_2 PSM1RW_3 PSM1RW_4 PSM1RW_5 PSM1RW_5 PSM1RW_6 PSM1RW_9 PSM1RW_9 PSM1RW_10 PSM1RW_10	CSM0 1 CSM0 3 CSM0 4 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 8 CSM0 8 CSM0 8 CSM0 10 CSM0 13 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 17 CSM0 17 CSM0 12 CSM0 12 CSM0 20 CSM0 21 CSM0 21 CSM0 22 CSM0 25	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 5 PSM1RW 6 PSM1RW 6 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 14	CSM0 1 CSM0 3 CSM0 4 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 6 PSM1RW 6 PSM1RW 7 PSM1RW 70 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM	CSMO 1 CSMO 3 CSMO 3 CSMO 4 CSMO 5 CSMO 6 CSMO 7 CSMO 7 CSMO 7 CSMO 10 CSMO 10 CSMO 11 CSMO 11 CSMO 11 CSMO 13 CSMO 14 CSMO 16 CSMO 16 CSMO 16 CSMO 18 CSMO 19 CSMO 17 CSMO 17 CSMO 12 CSMO 22 CSMO 22 CSMO 23	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1	CSMO_1 CSMO_3 CSMO_6 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_7 CSMO_10 CSMO_11 CSMO_11 CSMO_11 CSMO_11 CSMO_16 CSMO_16 CSMO_16 CSMO_16 CSMO_16 CSMO_12 CSMO_22 CSMO_22 CSMO_225	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 4 PSM1RW 5 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 12 PSM1RW 12 PSM1RW 12 PSM1RW 13 PSM1RW 12 PSM1RW 12 PSM1	CSM0_1 CSM0_3 CSM0_3 CSM0_4 CSM0_6 CSM0_6 CSM0_7 CSM0_7 CSM0_11 CSM0_11 CSM0_11 CSM0_13 CSM0_14 CSM0_14 CSM0_16 CSM0_16 CSM0_16 CSM0_21 CSM0_21 CSM0_21 CSM0_21 CSM0_21 CSM0_225	OUTGROUP
			PSM1RW_1 PSM1RW_2 PSM1RW_3 PSM1RW_4 PSM1RW_5 PSM1RW_5 PSM1RW_7 PSM1RW_10 PSM1RW_10 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_10 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_22 PSM1RW_	CSM0 1 CSM0 3 CSM0 4 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 8 CSM0 8 CSM0 10 CSM0 11 CSM0 13 CSM0 13 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 7 CSM0 20 CSM0 20 CSM0 21 CSM0 22 CSM0 22 CSM0 22 CSM0 22	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 4 PSM1RW 4 PSM1RW 5 PSM1RW 5 PSM1RW 5 PSM1RW 19 PSM1RW 19 PSM1RW 11 PSM1RW 10 PSM1RW 12 PSM1RW 22 PSM1RW 21 PSM1RW 22 PSM1RW 21 PSM1RW 22 PSM1RW 22 PSM1RW 20 PSM1RW 20 PSM	CSM0 1 CSM0 3 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CSM0 7 CSM0 7 CSM0 11 CSM0 13 CSM0 13 CSM0 13 CSM0 13 CSM0 14 CSM0 14 CSM0 16 CSM0 16 CSM0 16 CSM0 17 CSM0 12 CSM0 22 CSM0 22 CSM0 22 CSM0 22 CSM0 25	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 6 PSM1RW 6 PSM1RW 70 PSM1RW 70 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 12 PSM1RW 20 PSM1RW	CSM0 1 CSM0 3 CSM0 4 CSM0 5 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 3 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 10 PSM1RW 10 PSM1RW 20 PSM1RW 20 PSM1	CSMO_1 CSMO_3 CSMO_6 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_10 CSMO_11 CSMO_11 CSMO_11 CSMO_13 CSMO_14 CSMO_14 CSMO_14 CSMO_16 CSMO_16 CSMO_16 CSMO_12 CSMO_25 CSMO_22 CSMO_23 CSMO_23	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 4 PSM1RW 4 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 20 PSM1RW 20 PSM1	CSM0 1 CSM0 3 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CSM0 1 CSM0 11 CSM0 11 CSM0 13 CSM0 13 CSM0 14 CSM0 16 CSM0 16 CSM0 16 CSM0 20 CSM0 20 CSM0 21 CSM0 21 CSM0 21 CSM0 21 CSM0 22 CSM0 22	OUTGROUP
			PSM1RW_1 PSM1RW_2 PSM1RW_3 PSM1RW_4 PSM1RW_5 PSM1RW_5 PSM1RW_10 PSM1RW_10 PSM1RW_10 PSM1RW_112 PSM1RW_112 PSM1RW_113 PSM1RW_113 PSM1RW_114 PSM1RW_115 PSM1RW_115 PSM1RW_115 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_12 PSM1RW_15	CSM0 1 CSM0 3 CSM0 4 CSM0 4 CSM0 6 CSM0 7 CSM0 8 CSM0 8 CSM0 10 CSM0 11 CSM0 13 CSM0 13 CSM0 13 CSM0 13 CSM0 13 CSM0 14 CSM0 13 CSM0 14 CSM0 12 CSM0 20 CSM0 20 CSM0 21 CSM0 21 CSM0 22 CSM0 22 CSM0 22 CSM0 22 CSM0 22	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 2 PSM1RW 4 PSM1RW 5 PSM1RW 5 PSM1RW 5 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 22 PSM1RW 22 PSM1RW 22 PSM1RW 22 PSM1RW 22 PSM1RW 22 PSM1RW 26 PSM1RW 26	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 6 PSM1RW 6 PSM1RW 7 PSM1RW 7 PSM1RW 70 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 12 PSM1RW 22 PSM1RW 24 PSM1RW 24 PSM1RW 24 PSM1RW 25 PSM1RW 26 PSM1RW 2	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 3 PSM1RW 6 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 16 PSM1RW 16 PSM1RW 16 PSM1RW 17 PSM1RW 17 PSM1RW 17 PSM1RW 10 PSM1RW 20 PSM1RW 20	CSMO_1 CSMO_3 CSMO_6 CSMO_6 CSMO_6 CSMO_7 CSMO_7 CSMO_7 CSMO_10 CSMO_11 CSMO_11 CSMO_13 CSMO_14 CSMO_14 CSMO_14 CSMO_16 CSMO_16 CSMO_16 CSMO_16 CSMO_16 CSMO_12 CSMO_25 CSMO_22 CSMO_23 CSMO_23	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 4 PSM1RW 4 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 22 PSM1RW 22 PSM1RW 24 PSM1RW	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CSM0 7 CSM0 7 CSM0 11 CSM0 11 CSM0 13 CSM0 13 CSM0 14 CSM0 16 CSM0 16 CSM0 16 CSM0 20 CSM0 21 CSM0 21 CSM0 21 CSM0 21 CSM0 22 CSM0 22 CSM0 22	OUTGROUP
			PSM1RW_1 PSM1RW_2 PSM1RW_3 PSM1RW_4 PSM1RW_5 PSM1RW_5 PSM1RW_10 PSM1RW_10 PSM1RW_10 PSM1RW_11 PSM1RW_11 PSM1RW_11 PSM1RW_10 PSM1RW_10 PSM1RW_10 PSM1RW_10 PSM1RW_10 PSM1RW_20 PSM1RW	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 8 CSM0 8 CSM0 10 CSM0 11 CSM0 13 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 16 CSM0 20 CSM0 20 CSM0 20 CSM0 22 CSM0 25	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 2 PSM1RW 4 PSM1RW 4 PSM1RW 5 PSM1RW 5 PSM1RW 5 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 10 PSM1RW 22 PSM1RW 25 PSM1RW 25 PSM1RW 26 PSM1RW 10 PSM1RW 1	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 5 PSM1RW 6 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 11 PSM1RW 12 PSM1RW 22 PSM1RW 24 PSM1RW 24	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CS	OUTGROUP
			PSM1RW 1 PSM1RW 2 PSM1RW 3 PSM1RW 3 PSM1RW 6 PSM1RW 6 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 7 PSM1RW 10 PSM1RW 10 PSM1RW 11 PSM1RW 11 PSM1RW 15 PSM1RW 16 PSM1RW 16 PSM1RW 16 PSM1RW 17 PSM1RW 16 PSM1RW 16 PSM1RW 16 PSM1RW 22 PSM1RW 22 PSM1RW 24 PSM1RW 25 PSM1RW 24 PSM1RW 25 PSM1RW 24 PSM1RW 25 PSM1RW 26 PIBRW 16 PIBRW 16 PIBRW 17 PIBRW	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 7 CSM0 7 CSM0 1 CSM0 11 CSM0 11 CSM0 11 CSM0 11 CSM0 11 CSM0 13 CSM0 14 CSM0 16 CSM0 16 CSM0 20 CSM0 20 CSM0 20 CSM0 21 CSM0 21 CSM0 21 CSM0 21 CSM0 21 CSM0 21 CSM0 21 CSM0 22 CSM0 25	OUTGROUP
			PSMI RW, 1 PSMI RW, 2 PSMI RW 3 PSMI RW 4 PSMI RW 7 PIBRW 7 <td< td=""><td>CSM0.1 CSM0.4 CSM0.4 CSM0.6 CSM0.6 CSM0.7 CSM0.8 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.2 CS</td><td>OUTGROUP</td></td<>	CSM0.1 CSM0.4 CSM0.4 CSM0.6 CSM0.6 CSM0.7 CSM0.8 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.1 CSM0.2 CS	OUTGROUP
			PSM1RW_1 PSM1RW_2 PSM1RW_3 PSM1RW_4 PSM1RW_4 PSM1RW_5 PSM1RW_5 PSM1RW_10 PSM1RW_10 PSM1RW_10 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_112 PSM1RW_22 PSM1RW_22 PSM1RW_22 PSM1RW_22 PSM1RW_22 PSM1RW_22 PSM1RW_23 PTBRW_13 PTBRW_14 PTBRW_14 PTBRW_14 PTBRW_14 PTBRW_14 PTBRW_22 PSM1R	CSM0 1 CSM0 3 CSM0 4 CSM0 6 CSM0 6 CSM0 8 CSM0 8 CSM0 19 CSM0 13 CSM0 13 CSM0 13 CSM0 13 CSM0 13 CSM0 16 CSM0 16 CSM0 16 CSM0 20 CSM0 21 CSM0 21 CSM0 21 CSM0 22 CSM0 22 CSM0 24 CSM0 25	OUTGROUP

Figure 2. Gene trees for the ecotypes of *T. cristinae (top)* and the species pair *T. californicum and T. poppensis (bottom)* constructed for the nuclear locus 1 (N1). Green – *Ceanothus* ecotype; Blue – *Adenostoma* ecotype; Purple – *T. californicum*; Orange – *T. poppensis.*

HVC 1		00101001
HVC 2		
1100_2	— HVC_4	
HVC_5 HVC 6		
HVC_7		
HVC_9 HVC_10		
HVC_11		
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HVA_1		
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HVA_11 HVA_12		
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HVA_23 HVA_24		
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HVA_26		
LA_2		
LA_5		
LA 7		
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LA_18		
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LA_21		
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		 OUTGROUP
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PSM1RW_1		
PSM1RW 2 PSM1RW 2 PSM1RW 3		
CSMQ 2 PSM1RW_1 PSM1RW_2 PSM1RW_3 PSM1RW_4 PSM1RW_5		
CSM0_2 — PSMIRW 1 — PSMIRW 2 — PSMIRW 3 — PSMIRW 3 — PSMIRW 5 — PSMIRW 5 — PSMIRW 6 — PSMIRW 7		
CSMU2 1 PSMIRV 1 PSMIRV 1 PSMIRV 3 PSMIRV 3 PSMIRV 6 PSMIRV 6 PSMIRV 6 PSMIRV 6 PSMIRV 7		
CSMU_2 PSMTRW_1 PSMTRW_3 PSMTRW_4 PSMTRW_4 PSMTRW_6 PSMTRW 6 PSMTRW 6 PSMTRW 7 PSMTRW 7 PSMTRW 7 PSMTRW 10		
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Figure 3. Gene trees for the ecotypes of *T. cristinae (top)* and the species pair *T. californicum and T. poppensis (bottom)* constructed for the nuclear locus 2 (N2). Green – *Ceanothus* ecotype; Blue – *Adenostoma* ecotype; Purple – *T. californicum*; Orange – *T. poppensis.*





Figure 4. Gene trees for the ecotypes of *T. cristinae (top)* and the species pair *T. californicum and T. poppensis (bottom)* constructed for the nuclear locus 3 (N3). Green – *Ceanothus* ecotype; Blue – *Adenostoma* ecotype; Purple – *T. californicum*; Orange – *T. poppensis.*



Figure 5. Gene trees for the ecotypes of *T. cristinae (top)* and the species pair *T. californicum and T. poppensis (bottom)* constructed for the nuclear locus 4 (N4). Green – *Ceanothus* ecotype; Blue – *Adenostoma* ecotype; Purple – *T. californicum*; Orange – *T. poppensis.*



Figure 6. Gene trees for the ecotypes of *T. cristinae (top)* and the species pair *T. californicum and T. poppensis (bottom)* constructed for the mitochondrial locus CO1. Green – *Ceanothus* ecotype; Blue – *Adenostoma* ecotype; Purple – *T. californicum*; Orange – *T. poppensis.*

Population Structure:

Using the prior of K = 3, the program Structure strongly groups all species together. Notably, two individuals of *T. californicum* exhibit some genetic variation that is reflective of variation predominately observed in *T. poppensis*. Using the prior K = 6, the program Structure reflects the same grouping for *T. poppensis and T. californicum* as observed for K = 3, but attempts to break up the individuals of *T. cristinae* into multiple genetic clusters. Interestingly, the allopatric population of *T. cristinae* (LA) somewhat forms its own cluster (light blue). However, the ecotypes of *T. cristinae* nonetheless remain genetically mixed (*see Figure 7*).









Figure 7. Structure output for the number of predefined genetic clusters of K = 3 clusters and K = 6 clusters. Each individual is represented by a single bar. Each bar reflects the proportion of that individual's genotype (based on aggregated polymorphic site data), which are derived from any given genetic group. There were 254 total variable loci across all populations and all genes. The population individuals originally derived from are labeled below each plot: (1) HVC, (2) HVA, (3) LA, (4) CSMQ, (5) PSM1RW, (6) PTBRW.

Discussion

Past genetic analyses of speciation in *Timema* focused primarily on a single species, *T. cristinae* (Nosil, Egan, & Funk, 2008), and were based on genotypic or single gene DNA data (Law & Crespi, 2002), rather than multilocus genetic data. I presented here a multilocus genetic analysis of the genetic divergence within and between three species of *Timema* walking sticks. Variation was observed both among genes and among taxa. I first discuss patterns of variation within populations and then turn to the focus of the study: genetic divergence between populations and species. I conclude by highlighting particularly pressing avenues for future research.

Within-population genetic variation

Levels of within-population genetic diversity varied quite substantially both among genes and among the populations of *Timema* walking sticks examined in this study. In relation to the variation among genes, the high levels of genetic diversity in the mitochondrial gene CO1 are consistent with prior empirical findings in other study systems (Brown, et al., 1979; Lin & Danforth, 2004) and with the argument that mitochondrial DNA will tend to be more variable than nuclear DNA because mitochondrial DNA mutates faster and accumulates mutation faster (Brown, et al., 1979; Lin & Danforth, 2004). These data illustrate the root of genealogical discordance through differential variation across multiple loci and especially so between CO1 and the nuclear loci.

Between-population genetic differentiation

The differences between F_{ST} estimates at mitochondrial CO1 and nuclear genes as well as differences based on the geographic arrangement of populations reveal variable patterns of genetic differentiation and potential gene flow. The high levels of differentiation at CO1

between allopatric populations of *T. cristinae* compared with low levels of differentiation at CO1 between parapatric *T. cristinae* are likely due to two reasons: (1) the high rates of mitochondrial differentiation when gene flow is limited or absent, (2) in conjunction with low levels of differentiation between parapatric populations due to homogenizing gene flow (Nosil, et al., 2003). The lack of nuclear differentiation between both allopatric and parapatric pairs indicate that the ecotypes of *T. cristinae* are not highly differentiated, at least for the sample of nuclear genes examined in this study. Thus, genetic differentiation between phenotypically differentiated and partially-reproductively isolated ecotypes is likely restricted to a few genes, especially (or perhaps only) "speciation genes" subject to ecologically based divergent selection (Coyne & Orr, 2004; Nosil & Schluter, 2011; Wu, 2001). Consistent with this argument, previous research on heterogeneous genomic divergence among more than 500 AFLP loci detected outlier loci (using F_{ST}) whose genetic differentiation exceed neutral expectations (Nosil, et al., 2008). These outlier loci are likely affected by divergent selection and are thus driving the ecological speciation of *T. cristinae* and are the cause of initial reproductive isolation and assortative mating.

 F_{ST} estimates for *T. californicum* and *T. poppensis* indicate high levels of genetic differentiation between species at all nuclear loci in both parapatry and allopatry. Assuming that mitochondrial DNA diverges more quickly, one would expect equivalent or higher F_{ST} values for CO1 versus nuclear genes. However, we see lower F_{ST} at CO1 for *T. californicum* and *T. poppensis* in parapatry. This finding is highly suggestive of mitochondrial gene flow between these species; gene flow that results in reduced differentiation of the mitochondrial genome. As mentioned above, sequence data for CO1 from the allopatric population of *T. poppensis* and some individuals from the parapatric population of *T. poppensis* were unobtainable. This is

likely due to mutations in the priming region (since primers were developed from *T. cristinae* mRNA sequences) in most individuals from *T. poppensis.* However, since some sequences were obtainable for the parapatric population, and none for the allopatric population, it appears that there may be more differentiation in the allopatric population than the parapatric population, but these results remain inconclusive.

Gene trees and analyses of genetic structure both provide further support for our findings based on F_{ST}. Nuclear gene trees illustrate that species are more diverged than ecotypes. Also, population structure analyses with K=3 illustrate this point very clearly, strongly grouping individuals of the same species together. With K=6, we see the initiation of divergence between allopatric populations of *T. cristinae*. While previous research suggests ecologically based divergent selection as the driving force of genetic differentiation, past work shows that this process of ecological speciation can be constrained by gene flow and thus promoted by geographic separation (Nosil, 2007; Nosil & Crespi, 2004; Nosil, et al., 2008). The result of increased clustering of the allopatric population of *T. cristinae* is exaggerated by (1) the fact that mitochondrial DNA contributed the majority of polymorphic sites used to conduct the genetic structure analyses; (2) and the elevated rate of mitochondrial evolution. These two factors are indicated by the high F_{ST} value for the allopatric ecotype comparison (*Table 3*) and greater percent polymorphic site (%S) (Table 2). One final notable result is the appearance of two individuals from the population of *T. californicum* with mixed genetic structure (see Figure 7). This further suggests a marginal amount of gene flow between parapatric *T. californicum* and *T. poppensis*, also indicated by a low mitochondrial F_{ST} estimate. Additionally, one individual showed genetic characteristics of a hybrid, exhibited by the presence of haplotypes

from both populations. Future analyses using the program "NewHybrids" can test this explicitly (Anderson & Thompson, 2002).

Conclusions, Limitations and Future Directions

As one would expect from the general assumption of increased genetic differentiation as speciation unfolds, we found ecotypes to be significantly less differentiated than species. Interestingly however, taxon pairs subject to geographic isolation (i.e. allopatry) did not predict higher levels of nuclear differentiation than their parapatric counterparts (for both species and ecotypes). This finding supports the hypothesis that ecologically based divergent selection is the key driver of genetic differentiation between host-plant associated ecotypes of *Timema cristinae* and also that selection is only acting on a few speciation genes. Additionally, the mitochondrial gene CO1 appears to be "flowing" readily in parapatry for diverging ecotypes and recently diverged species. This finding is not uncommon. Ferris et al. (1983) found mitochondrial DNA flowing between different species of mice and comments that, if this is common, it may require a redefinition of a biological species to specify the use of only nuclear genes. Nearly 30 years have passed since the latter paper was published and despite the fact that multiple other studies have demonstrated the same phenomenon, no redefinition of a biological species has been made, much less the formation of a single cohesive and all inclusive definition of a "species".

Genealogical discordance was observed here, especially between mitochondrial and nuclear gene tress, thus emphasizing the importance of multilocus genetic data for

phylogenetic purposes. While the species level mitochondrial gene tree is consistent with the finding of Law and Crespi (2002), which indicates a mixed phylogenetic relationship between *T. califronicum* and *T. poppensis*, the inclusion of four nuclear genes provides sufficient evidence for the separation of these taxa as distinct species. The results for CO1 are consistent with the general patterns observed in animal taxa. Funk and Omland (2003) reviewed studies of mitochondrial DNA divergence using 2319 assayed species and found that 23 percent of examined *distinct species pairs* were not reciprocally monophyletic based on mitochondrial DNA. Overall, these results indicate that genetic studies relying on a single locus may result in misleading conclusions.

Further analyses with the taxa employed here are necessary for clarifying our understanding of ecological speciation. Multilocus analyses with a larger sample from populations of *T. cristinae* are requisite for more precisely investigating how much genetic differentiation is reflected at neutral loci. Although genetic differentiation may be minimal at these loci, obtaining significant F_{ST} estimates will provide important information about background differentiation along the speciation continuum for genes not directly involved in initial divergence but that may play a role in the eventual complete reproductive isolation of two diverging populations. Additionally, statistically based estimates of gene flow need to be made using "Isolation with Migration" models (Nielsen & Wakeley, 2001). While F_{ST} can provide useful information about genetic divergence, it does not allow us to differentiate between recent divergence, gene flow or some combination of the two. Another important direction for further research is the development of primers for CO1 based on *T. poppensis*. The unsuccessful sequencing of CO1 in most individuals of *T. poppensis* confers the need for new primers, which would allow for complete analyses across all populations and all genes.

Finally, although it is a lofty goal, we need to reach a better solution to the "species problem." The use of a singular working definition of a "species" would immensely benefit the fields of evolutionary biology, ecology, and conservation (Mallet, 2007) and would especially be useful when studying evolutionary divergence and how species are formed. The most common approach to a solution is to aggregate many of the working concepts of a species (Mallet, 1995). For example, one could use information about morphology, ecological role, and amount of genomic differentiation to define a species. However, all of these realms of distinction require a fairly arbitrary line to be drawn. How different do species have to look, or behave? How genetically differentiated must species be? After conducting this study, it is clear to me why the definition of a species is so unclear, especially on the genetic level. Depending on where in the genome one looks, or what genome one looks at, varying levels of genetic differentiation will likely be observed. However, by conducting studies using multilocus nuclear data (on the order of hundreds or thousands of loci) across a wide range of taxa, we may be able to more precisely define where to draw a line of distinction based on genetic differentiation. This new definition should take into consideration key factors such as ecological roles and general morphological difference, but fundamentally, genes are driving the processes of speciation and an extensive survey of genetic differentiation will ultimately contribute the most information to our understanding of evolutionary divergence and thus how to define a "species".

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